Comparative Anatomy of Lachnanthes and Lophiola (Haemodoraceae)1)

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Summary

Aspects of vegetative and floral anatomy of the monotypic genera *Lachnanthes* and *Lophiola* are described and compared. *Lophiola* and *Lachnanthes* are anatomically very distinct from one another and the taxonomic placement of both genera within the same tribe is not supported. The relationships of the genera to other taxa remain unclear.

Introduction

The Haemodoraceae is a relatively small family of monocotyledons. The most recent treatment by Geering (1969) circumscribed the family as composed of rhizomatous, often stoloniferous, scapose to subscapose herbs with basal, equitant, "Iris-like" leaves which are basally sheathing and distally unifacial by longitudinal folding and fusion. The inflorescence is terminal and usually branched and is composed of bisexual, often externally tomentose flowers with 6 tepals, 3—6 stamens, and a superior to inferior ovary; fruits are usually loculicidal capsules with endospermous seeds.

The family (sensu Geerinck 1968, 1969) contains 13 genera and 76 species, with distributions in South Africa, Australia, South America, Central America, Mexico, and eastern North America. However, a precise delimitation and satisfactory intrafamilial classification of the Haemodoraceae remain unavailable. Several authors have proposed different combinations of tribes and genera (see Geerinck 1968: ROBERTSON 1976). Among recent family treatments, Melchior (1964), in the "Syllabus der Pflanzenfamilien", recognized three tribes: Haemodoreae, Conostylideae, and Conanthereae. Hutchinson (1934, 1959, 1973) grouped the Haemodoraceae with five other families in the order Haemodorales and divided the family into two tribes: Haemodoreae with 11 genera and Conostylideae with 6 genera; the Conanthereae being included with the family Tecophilaeaceae of the Liliales. Geerinck (1969) also recognized the Haemodoreae and Conostylideae although differed from Hutchinson by removing Lanaria from the family, transferring Lophiola to the Haemodoreae, and merging genera previously recognized as distinct. Among contemporary system makers, Cronquist (1968) classified the Haemodoraceae within the Liliales of the Liliidae. TAKHTAJAN'S (1969) treatment essentially agreed with that of Cronquist, and regarded the Hypoxidaceae and Velloziaceae as closely related families. Thorne

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(1976) lowered the family to subfamilial status within the *Liliaceae* and Dahlgren (1975, 1977) placed the *Haemodoraceae*, along with the *Pontederiaceae* and *Philydraceae*, in the *Haemodorales* of the superorder *Lilianae*.

The eastern North American representation of the family consists of two superficially similar and apparently monotypic genera, Lachnanthes caroliniana (Lam.) Dandy and Lophiola aurea Ker-Gawler. Although some recent publications still refer to the species of Lophiola as L. americana (Pursh) Wood, Robertson (1976) presented a convincing case that the name L. aurea Ker-Gawler has priority. Lachnanthes caroliniana is locally common and ranges, with several disjunctions, from southern Nova Scotia to Cuba, including the Atlantic and Gulf coastal plain of the southeastern United States. Lophiola aurea is somewhat rarer, occurring as a disjunct along the coastal plain in western Nova Scotia, the pine barrens of New Jersey, and in scattered localities from southeastern North Carolina to the Florida panhandle and southern Mississippi. Both species typically grow in bogs and low, wet areas in savannahs and pinelands, although Lachnanthes caroliniana is also common in swamps and roadside ditches.

The taxonomic position of Lachnanthes within the Haemodoraceae is apparently reasonably well established. Robertson (1976), who provided a thorough morphological characterization of the two genera, noted that, in his opinion, Lachnanthes clearly belongs to the tribe Haemodoreae, being most closely related to Haemodorum of Oceania and Dilatris of South Africa. The evolutionary relationships of Lophiola, however, are quite unclear, having been placed variously in the Amaryllidaceae, Hypoxidaceae, and Haemodoraceae (Geerinck 1969).

No detailed anatomical, embryological, or palynological studies have been undertaken on either *Lachnanthes* or *Lophiola*. Accordingly, this initial anatomical investigation was undertaken to provide information which may eventually provide a more definitive family circumscription and intrafamilial classification.

Materials and Methods

Material was collected in the field and voucher specimens deposited in the herbarium of The University of North Carolina at Chapel Hill (NCU). Vegetative organs, floral buds, and mature flowers were fixed in formalin-acetic acid-alcohol (FAA). Both fresh sections and prepared slides were employed for study. Permanent slides were prepared using standard anatomical techniques of dehydration, paraffin infiltration, and microtome sectioning (Johansen 1940). Sections were stained with safranin-fast green or safranin-haematoxylin. Both transverse and longitudinal sections of roots, rhizomes, scapes, and leaves were made. Tepal vestiture was examined from temporary wet mount slides. Floral vasculature was studied utilizing a combination of serial sections of mature floral buds and cleared flowers. Mature whole flowers were cleared in 5.25 % sodium hypochlorite and stained with pararosaniline hydrochloride (Boke 1970). Stomatal ontogeny was observed from temporary whole mounts of immature leaf bases, fixed in 3:1 absolute ethanol and glacial acetic acid (CARNOY'S Fluid) and stained with acetocarmine (TOMLINSON 1974). Vessels were studied from both longitudinal sections and macerations; thin slivers of fresh roots, rhizomes, and leaves were macerated in 10% aqueous chromic acid: 10% aqueous nitric acid (Jeffrey's solution) for 2-3 days, teased gently with needles, and mounted unstained in HOYER'S solution. Prepared slides are deposited in the anatomical slide collection of The University of North Carolina at Chapel Hill.

Observations

Vegetative Anatomy

Lachnanthes caroliniana

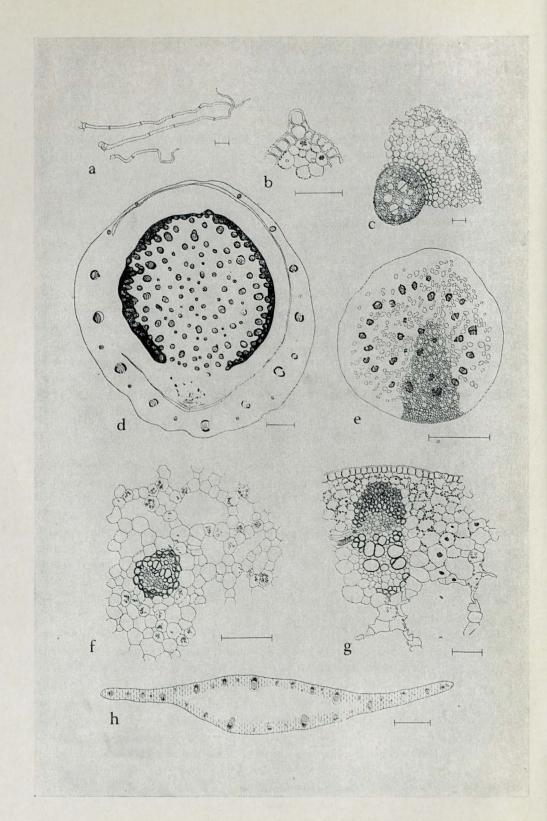
Root (Fig. 1c). Epidermis uniseriate, with longitudinally extended thin-walled cells. Cortex with a peripheral, uniseriate exodermal layer of crushed cuboidal cells, an extensive aerenchymatous middle zone, and a 2—3 layered inner zone of thick-walled, sclerified cells; internal cortical cells radially aligned with endodermal cells. Endodermis uniseriate, with thick-walled cells which are uniformly lignified on inner and outer tangential walls. Pericycle uniseriate, slightly lignified. Xylem polyarch, with 10—11 protoxylem poles and a central core of sclerified parenchyma; vessels with simple perforation plates (Fig. 6a, b), ca. 0.8—1.4 mm long. Phloem in clusters of 1—2 sieve tube elements flanked by smaller elements and companion cells, alternating with protoxylem poles. Crystals absent.

Rhizome (Fig. 1e, f). Epidermis uniseriate; cells thin-walled, longitudinally extended; outer wall thickly cutinized with tuberculate outer surface. Ground tissue differentiated into narrow outer region of thin-walled, spheroidal parenchyma and extensive inner region of schizogenous aerenchyma, forming an alveolar network. Endodermis absent. Vasculature atactostelic; bundles radially extended, enclosed by 1—2 layers of sclerenchyma. Xylem endarch; vessels to ca. 1.9 mm long, with multi-barred perforation plates (Fig. 6c). Phloem oriented in a wide tangential band. Starch grains numerous in aerenchyma. Raphides present, scattered throughout ground tissue.

Rhizome at aerial shoot base considerably expanded (Fig. 1d). Intercellular spaces small. Stelar region enclosed by a multilayered sclerenchyma sheath in endodermal region; no distinct endodermis present. Vascular bundles collateral to amphivasal. Xylem typically C-shaped, enclosing phloem.

Scape. Epidermis uniseriate; cells rectangular to cuboidal in transection, with thick, warty outer cuticles. Cortex of an outer chlorenchymatous zone and inner aerenchymatous zone separated by a ring of fibers; dark-staining, rod-shaped bodies present in chlorophyllous cells. Vascular bundles collateral, scattered throughout central region; bundles wholly or partially enclosed by sclerenchymatous sheath. Starch grains and crystals absent.

Leaf (Fig. 1g, h). Leaves "Iris-like", bifacial and sheathing at base, unifacial above by longitudinal fusion. Leaves narrowly spindle shaped in transection, with two marginal flanges. *Epidermis* thin-walled; cells cubic to oblong in transection, outer walls with a thick, often papillate or ridged cuticle. *Stomata* with two laterally flanking subsidiary cells (Fig. 5e); guard cells each with two cutinized ridges, projecting into stomatal openings; substomatal cavities small (Fig. 1g). *Stomatal development* initiated by an unequal cellular division of a protodermal cell at its distal end (toward leaf apex, Fig. 5a), the resultant guard cell mother cells surrounded by four "neighboring cells" (sensu Tomlinson 1974) (Fig. 5b); the two lateral neighboring cells divide unequally and non-obliquely to form two lateral "contact cells"



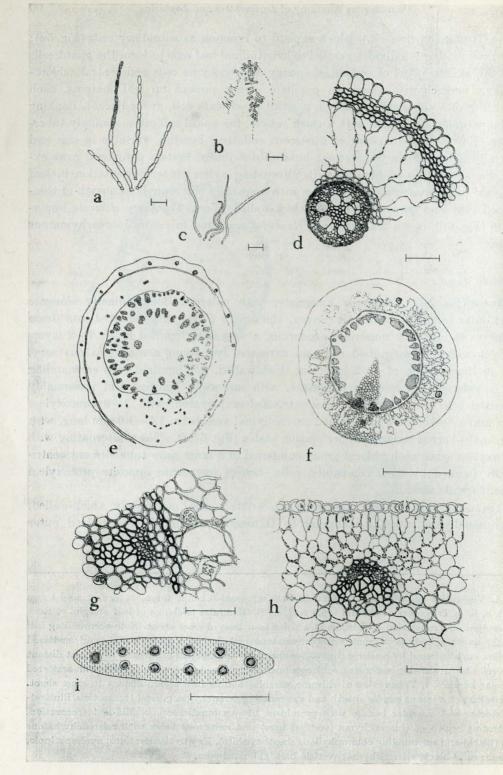
(sensu Tomlinson, loc. cit.), which expand to function as subsidiary cells (Fig. 5c); guard cell mother cell divides parallel to longitudinal leaf axis to form the guard cells (Fig. 5d, e). Mesophyll of peripheral spongy chlorenchyma cells and central achlorophyllous aerenchyma, occasional papillate glands present on leaf margins, each consisting of a spherical apical cell, a small basal stalk cell, and adjacent, flanking epidermal cells; gland cells with a thick cuticle, the apical cell often minutely tuberculate. Vasculature composed of numerous collateral bundles, variable in size and generally ellipsoidal in transection; bundles distributed in two peripheral rows except for single files of bundles with alternating xylem/phloem orientation in leaf marginal extensions; vascular bundles with a sparsely chlorophyllous sheath of thin-walled cells and prominent cap of thick-walled fibers. Tracheary elements imperforate (Fig. 6d), up to 2.1 mm long. Raphides scattered in central aerenchymatous tissue.

Lophiola aurea

Root (Fig. 2d). Epidermis uniseriate, with enlarged, longitudinally elongate cells; outer tangential cell wall convex, thickened. Cortex differentiated into (from periphera to center) a uniseriate exodermis, a uniseriate lignified layer, 1–2 layers of thick-walled, non-lignified cells, an extensive lysigenous aerenchymatous zone, and an inner region of 1–2 layers of thick-walled, non-lignified cells surrounding the endodermis. Endodermis uniseriate, with uniformly lignified and presumably suberized cell walls. Pericycle uniseriate. Xylem polyarch, with 7–9 protoxylem poles and a central core of sclerified parenchyma; vessels ca. 1.5–1.9 mm long, with long, multi-barred scalariform perforation plates (Fig. 6e, f). Phloem alternating with protoxylem arms, each phloem group composed of a large sieve tube element centripetally flanked by 3–5 companion cells. Lateral roots arise opposite protoxylem poles. Crystals absent.

Rhizome (Fig. 2f, g). Epidermis uniseriate; cells moderately thick-walled, longitudinally extended, rectangular in transection, with thinly cutinized outer

Fig. 1. Vegetative anatomy of Lachnanthes. Bars represent 0.1 mm for a, b, c, f, g and 1 mm for d, e, h. a, Trichomes of abaxial tepal surface. b, Glandular trichome of leaf margin. c, Root transection. d, Transection of rhizome at a point near base of erect shoot. Note surrounding leaf sheath, axillary bud, prominent sclerenchymatous endodermal region (black), and scattered bundles, each enclosed by a sclerenchymatous sheath. e, Transection of rhizome at a point distant to erect shoot, showing extensive schizogenous aerenchymatous ground tissue and scattered vascular bundles. f, Transection of rhizome vascular bundle in a region distant to erect shoot. Note sclerenchymatous bundle sheath and surrounding aerenchyma. g, Leaf transection, illustrating stoma, and vascular bundle with prominent fibrous bundle cap. h, Mid-leaf transection, illustrating region of chlorenchyma (vertical bars) and numerous veins with sclerenchymatous caps (black) and surrounding chlorophyllous sheaths (white). Key to tissues: lines, xyelm; stipple, phloem; solid black, sclerenchyma; verticle bars, chlorenchyma.



walls. Cortex differentiated into a 2-4 layered, sclerified, subepidermal region with sparse intercellular spaces and an inner unlignified lysigenous aerenchymatous zone. Endodermis uniseriate, highly lignified. Pericyclic region multi-layered, partially sclerified. Vascular bundles collateral, in a single ring. Xylem endarch; tracheary elements presumably imperforate (Fig. 6g), up to 1.4 mm long. Pith composed of thick-walled, unlignified cells with prominent intercellular spaces. Starch grains numerous in ground tissue. Raphides scattered in internal cortical zone.

Rhizome at base of aerial shoot (Fig. 2e) enlarged and anatomically altered. Cortex devoid of aerenchyma, with few starch grains. Vasculature atactostelic, with numerous outer, collateral bundles and fewer inner, amphivasal bundles; sheath cells of bundles occasionally smaller, with numerous starch grains. Ground tissue centrally partially sclerified, with sparse intercellular spaces and numerous starch grains.

Scape. Epidermal cells longitudinally elongate, with thick inner and outer tangential walls, thinly cutinized. Stomata numerous, resembling those of the leaf. Ground tissue differentiated into a narrow outer chlorenchymatous zone of spherical, thin-walled cells with prominent intercellular spaces and an extensive inner parenchymatous zone of outer thick-walled and central thin-walled cells. Vascular bundles collateral, scattered in periphery of inner parenchymatous zone; inner bundles with a C-shaped region of tracheary elements partially to wholly encircling the phloem and an outer fibrous bundle cap usually dividing the phloem into two or more discrete units; bundles near periphery with xylem and phloem positioned along a tangential plane; outermost bundles (leaf or tract traces) with a prominent sclerenchymatous outer cap and radially extending fibers which separate the phloem into two groups. Starch grains essentially absent. Raphides scattered in inner region of ground tissue.

Leaf (Fig. 2h, i). Leaves basally bifacial and sheathing, distally unifacial by longitudinal fusion, oriented perpendicular to the scape. Leaves narrowly-elliptic to narrowly oblong in transection. *Epidermis* uniseriate, cells longitudinally elongate, cubic to radially oblong in transection; inner and outer tangential walls thick; cu-

Fig. 2. Vegetative anatomy of Lophiola. Bars represent 0.1 mm for a, b, c, d, g, h and 1 mm for e, f, i. a, Trichomes of adaxial tepal surface. b, Trichomes of apical, adaxial tepal surface. c, Trichomes of abaxial tepal surface. d, Transection of root, showing large epidermal cells and lysigenous aerenchymatous cortex. e, Transection of rhizome at a point near base of erect shoot, showing surrounding leaf sheath, endodermis, and peripherally scattered, concentric, vascular bundles. f, Transection of rhizome at a point distant to erect shoot, illustrating lysigenous aerenchymatous inner cortex, endodermis, and ring of collateral vascular bundles. g, Transection of rhizome vascular bundle at a point distant to erect shoot. h, Leaf transection, illustrating stomata, mesophyll layers, and vascular bundles i, Leaf transection, showing chlorenchymatous region (vertical bars) and veins. Each vein is surrounded by an inner sclerenchymatous sheath (black) and an outer chlorophyllous sheath (white). Key to tissues: lines, xylem; stipple, phloem; solid black, sclerenchyma; vertical bars, chlorenchyma.

ticle moderately thick. Stomata of two longitudinally oriented guard cells surrounded by four cruciately arranged "neighboring cells", none of which are specialized as subsidiaries (Fig. 5i): guard cells with two cutinized ridges of wall material which project into stomatal cavity (Fig. 2h); stomatal cavities small. Stomate development initiated by an unequal distal division of a protodermal cell, the smaller, distal product functioning as the guard cell mother cell (Fig. 5f, g); cells undergo some expansion before the guard cell mother cell divides longitudinally into two guard cells (Fig. 5h). Mesophyll composed of an outer region of two or three layers of loosely arranged, columnar palisade cells and a central region of spheroidal spongy chlorenchyma cells, which are usually crushed and achlorophyllous. Vasculature consisting of an ellipse of vein bundles positioned just inside the palisade cells; a single vein, or two partially fused veins, occur at each leaf margin; bundles uniform, collateral, orbicular in transection, surrounded by an outher sheath of thin-walled, sparsely chlorophyllous cells and an inner sheath of thick-walled, sclerenchyma cells. Xylem endarch, with occasional protoxylem lacunae; tracheary elements presumably imperforate, ranging to 2.3 mm in length (Fig. 6h). Phloem characteristically separated into two groups by fibers extending radially from the inner bundle sheath to the outer metaxylem. Raphides scattered throughout achlorophyllous central region.

Floral Anatomy

Lachnanthes caroliniana

General morphology and anatomy. Flowers are bisexual, actinomorphic, bracteate, and pedicellate with a perianth of six epigynously positioned tepals in two whorls: an outer whorl of three linear-triangular tepals and an inner whorl of three larger oblanceolate tepals. The androecium consists of a single whorl of three stamens positioned opposite the inner, larger, somewhat involute tepals. The stamens are exserted, and bear basifixed, bilocular anthers with longitudinal introrse dehiscence. The gynoecium consists of a syncarpous, tricarpellate, pistil with an inferior, three-lobed ovary. Each of the three locules contains a transversely oriented, protruding, peltate, axile placenta bearing 9–12 bitegmic, anatropus ovules. The ovules are positioned along the entire margin of the peltate placenta with their micropyles oriented toward the juncture of the placental stalk and central ovary axis and are, thus, pleurotropous, in position with the raphe facing the center of each peltate placenta. The single, terete style has a slightly expanded, unlobed stigma.

A whitish tomentum covers the inflorescence branches, pedicles, outer ovary wall, and abaxial tepal and bract surfaces. Trichomes are simple (unbranched), uniseriate, and composed of a short basal cell and 2—8 (usually four) elongate and filiform cells, including a tapering apical cell (Fig. 1a). The adaxial tepal surfaces are glabrous.

Raphides are scattered in all parts of the flower, occurring most profusely in the placentae.

Vascular anatomy. The pedicel contains an irregular complex of vascular tissue, consisting of approximately 9-12 concentrically arranged peripheral bundles and

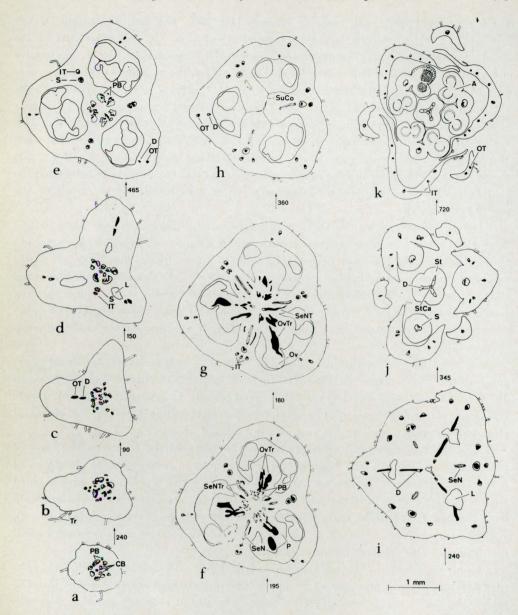


Fig. 3. Lachnanthes. Camera lucida drawings of serial transections of mature bud from pedicel (a) to apex (k). Distance between successive sections indicated in microns. Key to tissues: solid black, xylem or lateral trace; stipple, phloem. Abbreviations: A, Anther; CB, Central Bundles; D, Dorsal carpellary bundle; IT, Inner Tepallary bundle; L, Locule; OT, Outer Tepallary bundle; Ov, Ovule; OvTr, Ovule Trace; P, Placenta; PB, Placental Bundle; S, Stamen bundle; SeN, Septal Nectary; SeNTr, Septal Nectary Trace; St, Style; StCa, Stylar Canal; SuCo, Sutural Commissure; Tr, Trichome.

three centrally positioned vascular strands (Fig. 3a). The peripheral strands are collateral in organization and have a normal orientation, i.e., xylem positioned abaxially, phloem adaxially, whereas the three smaller central bundles are generally inverted. As the pedicel enlarges to form the receptacle, the vasculature becomes more profusely and irregularly branched (Fig. 3b). At the level of the receptacle base, three radially aligned pairs of collateral bundles extend from the central vascular complex (Fig. 3c). There three pairs of bundles traverse the outer ovary wall, with each pair branching at a higher level into two or three tangentially oriented strands prior to entering the outermost whorl of tepals; further branching in the outer tepals rarely occurs. The inner bundle of each pair functions as the dorsal carpellary bundle and extends the entire length of the carpel wall, terminating at the apex of the style (Fig. 3d-k).

At the base of the locules three additional radially aligned bundle pairs extend from the central complex and traverse the outer ovary wall in a position opposite the septa (Fig. 3d). The more peripheral bundle of a pair divides at a level just above placenta insertion into three collateral bundles which subsequently enter the inner whorl of tepals; additional branching of the vasculature within each inner tepal may result in as many as nine tepal traces. The inner bundle pairs which are located opposite the septa are amphicribral or collateral and supply the vasculature to the stamens (Fig. 3e—k).

At a level just below the placentae, the central complex of vascular tissue is organized into three diffuse and dissected pairs of "placental" bundles (sensu Sterling 1972), each pair positioned centripetal and opposite to a carpel (Fig. 3e). Ovule traces arise from the pair of placental bundles and radiate through the placental stalk to vascularize the circularly arranged marginal ovules (Fig. 3f, g).

Three "septal nectaries" are initiated in the septa of the ovary at the level of placenta insertion (Fig. 3f). Each nectary, as viewed in transection, is composed of a radially elongate schizogenous canal that parallels the septum wall and is lined with a single layer of differentially staining epithelial cells. Lateral traces originate from the placental bundles and extend to either side of the septal nectary (Fig. 3f—g). Each nectary traverses the entire verticle length of the septum and opens by a small distal pore at the apex of the ovary (Fig. 3i).

At an upper level in the ovary, three ventral sutural commissures (Fig. 3h) connect the locules with the base of an open, three-lobed canal which extends into the style. The stylar canal is composed of loosely arranged cells throughout its length and a small central lacuna. No ventral sutures are evident in the style (Fig. 3i—k).

Lophiola aurea

General morphology and anatomy. The flowers of *Lophiola* are bisexual and actinomorphic, with bracteate pedicels. The perianth is composed of six similar lanceolate to narrowly triangular tepals positioned in two whorls. The tepals are epihypogynous in position, arising from the ovary at about two-thirds the ovary height. The androecium is composed of two whorls of three stamens each, the outer whorl

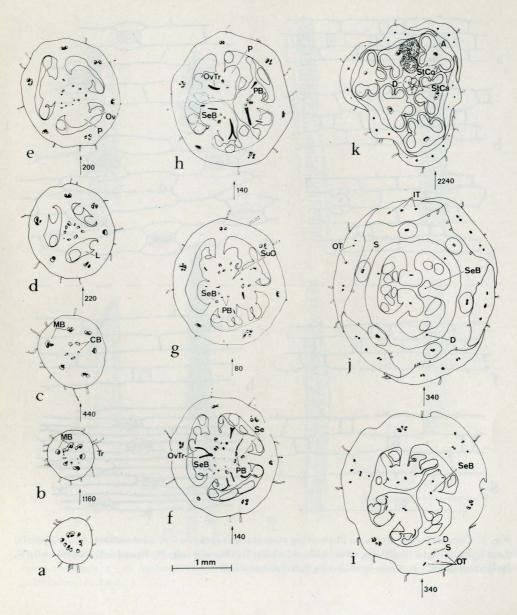


Fig. 4. Lophiola. Camera lucida drawings of serial transections of mature bud from pedicel (a) to apex (k). Distance between successive sections indicated in microns. Key to tissues: solid black, xylem or lateral trace; stipple, phloem. Abbreviations: A, Anther; CB, Central Bundle; D, Dorsal carpellary bundle; IT, Inner Tepallary bundle; L, Locule; MB, Major Bundle; OT, Outer Tepallary bundle; Ov, Ovule; OvTr, Ovule Trace; P, Placenta; PB, Placental Bundle; S, Stamen bundle; SeB, Septal Bundle; StCa, Stylar Canal; StCo, Stylar Commissure; SuO, Sutural Opening, Tr, Trichome.

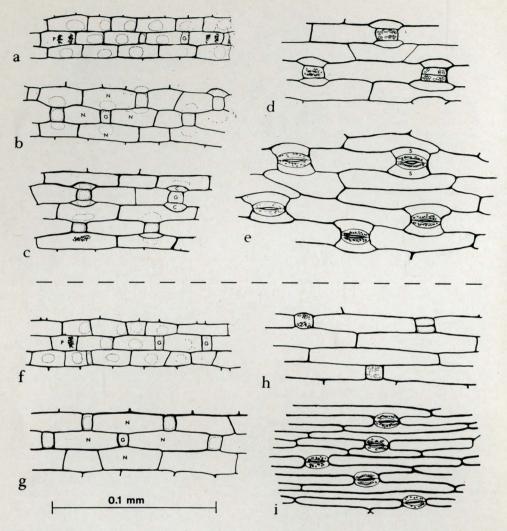


Fig. 5. Camera lucida diagrams illustrating stomatal ontogeny. a—e, Lachnanthes; f—i, Lophiola. Leaf apex to right in all diagrams. Abbreviations: C, Contact cell; G, Guard cell mother cell; N, Neighboring cell; P, Protodermal cell; S, Subsidiary cell.

situated opposite the outermost whorl of tepals. Anthers are basifixed and bilocular at maturity with longitudinal, introrse dehiscence. The gynoecium is syncarpous and tricarpellate. The half-inferior ovary is 3-locular with axile placentation below and unilocular with three parietal placentae above. The placentae are slightly protruding, each bearing numerous (ca. 30) ovules. Ovules are anatropous, bitegmic, and heterotropous, i.e., have a variable micropyle orientation with respect to the ovary. The single style is terete and the stigma unlobed.

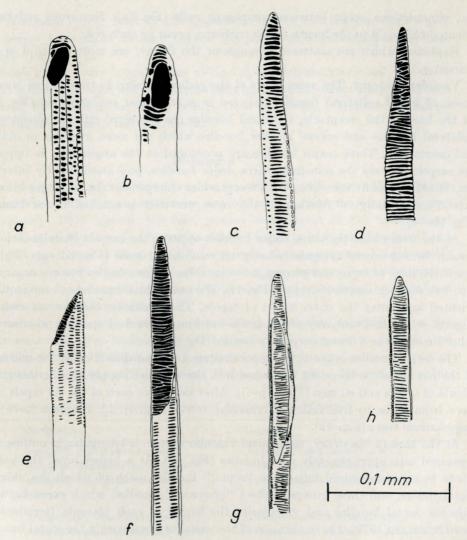


Fig. 6. Camera lucida drawings of tracheary elements, including perforation plates of obvious vessel elements. a—d, *Lachnanthes*. a, b, Root. c, Rhizome. d, Leaf. e—h, *Lophiola*. e, f, Root. g, Rhizome. h, Leaf.

Inflorescence branches, pedicels, hypanthium, and abaxial surfaces of the bracts and tepals are covered with a dense whitish tomentum. The whitish colored trichomes are simple (unbranched), uniseriate, and bicellular. The basal cell of each trichome is somewhat isodiametric, whereas the apical cell is long and filiform (Fig. 2c). Numerous short, terete, unicellular trichomes with rounded ends are found at the apex of each tepal, occurring along the extreme apical margins and midvein on the adaxial surface (Fig. 2b). A dense tuft of bright orange trichomes occurs at the base of the adaxial surface of each tepal; these hairs are simple, uniseriate, multicellular, and moniliform,

i.e., constrictions occur between component cells (Fig. 2a). Numerous striations, oriented obliquely to the length of the trichome, occur on each cell.

Raphides, which are scattered throughout the flower, are most plentiful in the placentae.

Vascular anatomy. The vasculature of the pedicel, as seen in transection, is composed of 9-12 collateral bundles oriented in a triangular configuration (Fig. 4a). At the base of the receptacle, the floral bundles are organized into six conspicuous collateral bundles and several smaller bundles which are more irregular in outline and orientation. Three major bundles are positioned at the angles and lie opposite the carpels whereas the remaining three major bundles, positioned slightly internal, are oriented opposite the septa. With receptacular enlargement the six major bundles extend peripherally in relation to the more centrally positioned minor bundles (Fig. $4\,\mathrm{b}-\mathrm{e}$).

At the ovary base the three major bundles opposite the carpels characteristically branch into 2-several irregularly oriented vascular strands (Fig. 4d-g). Slightly below the level of tepal and stamen insertion these major bundles become organized into five distinct vascular strands (Fig. 4i). The outer three bundles are tangentially oriented and enter the outer whorl of tepals. The remaining bundles are radially aligned, with one eventually entering the outermost whorl of stamens whereas the other functions as a dorsal carpellary bundle (Fig. 4j, k).

The major bundles opposite the septa undergo irregular dissection at the mid-level of the ovary before becoming organized into the vascular supply for the innermost whorls of tepals and stamen (Fig. 4g—j). After initiation, each of the six tepals may have from three to five collateral vascular traces (Fig. 4j, k). Stamens have one amphicribral trace (Fig. 4k).

At the base of the ovary, the central vascular system is triangular in outline and organized into approximately nine bundles (Fig. 4d). At a higher level, the vasculature becomes organized into three "septal" bundles, each of which lies along a septal radius, and three groups of 2—4 "placental" bundles, which occur between adjacent septal bundles and vascularize the ovules of each placenta (terminology sensu Sterling 1972). The orientation of the vascular tissue within the septal bundles is variable, being collateral, bicollateral, or collateral with phloem and xylem oriented tangentially. The placental bundles are almost always inverted with phloem centripetal to the xylem. Slightly above the ovary base, each placental bundle branches with the resulting veins terminating within two (rarely three) ovules. Septal bundles remain unbranched as they traverse the central ovary, only rarely contributing a vascular trace to the placenta (Fig. 4f—i).

At about the level of tepal and stamen attachment, sutural openings radially divide each placenta, resulting in a apically unilocular ovary. Two placental bundles (each derived from the originally axile placentae of two adjacent carpels) and a single septal bundle vertically traverse the now parietal placentae. Each placental bundle develops lateral branches which subsequently bifurcate, regularly vascularizing two of the four ranks of ovules occurring on the parietal placentae. The placental

bundles terminate at the level of the uppermost ovules whereas the three septal bundles persist for a short distance beyond the placentae, terminating at the base of the style (Fig. 4g—j).

The locule of the ovary is continuous with an open 3-lobed stylar canal. The outer epidermal cells of the style are characteristically papillose in shape. Each canal lobe is traversed by a continuous dorsal bundle from the ovary. Sutural commissures delineate the three canal lobes (Fig. 4j-k).

Discussion

From the comparison in Table 1, it can be seen that the two eastern United States representatives of the *Haemodoraceae* are anatomically distinct despite their superficial similarity. Both species, however, possess aerenchyma in the underground stems, a reflection of the generally wet to flooded habitats in which they grow, and raphides in vegetative and floral parts. Interestingly, the aerenchyma in *Lachnanthes* is schizogenous in origin whereas that of *Lophiola* appears predominantly lysigenous in development. Leaf vascular bundles of both plants are surrounded by ill-defined chlorenchymatous bundle sheaths.

Unfortunately, the general paucity of published anatomical information on the *Haemodoraceae* and allies makes intra- and interfamilial comparisons difficult and premature. Schulze (1893) provided a superficial, though significant, synopsis of the comparative anatomy of the *Liliaceae*, *Haemodoraceae* (tribe *Haemodoreae* only),

Table 1. Comparison of major anatomical features of Lachnanthes and Lophiola

	Lachnanthes	Lophiola
Root	Polyarch; vessel elements with simple perforations; inner cortex sclerified; epidermal cells not enlarged	Polyarch; vessel elements with scalariform perforations; cortex aerenchymatous; epidermal cells enlarged
Rhizome	Atactostelic; vessel elements with scalariform perforations; endodermis absent; ground tissue aerenchymatous	Vascular bundles in single ring; vessels absent; endodermis present; cortex aerenchymatous
Leaf	Vascular bundles numerous, variable in size and ellipsoidal in transection, with sclerenchymatous caps; vessels absent; palisade cells absent; subsidiary cells present; raphides present	Vascular bundles 8-12, of uniform size and circular in transection, with sclerenchymatous sheaths; vessels absent; palisade layers present, subsidiary cells absent; raphides present
Flower	Placental bundles 2 per carpel; septal bundles absent; septal nectaries present; septal commissures present; stylar commissures absent; ovules anatropous, bitegmic, pleurotropous; raphides present	Placental bundles 2-4 per carpel; septal bundles present; septal nectaries absent; septal opening present; stylar commissure present; ovules anatropous, bitegmic, heterotropous; raphides present

Hypoxidoideae of the Amaryllidaceae (including the tribe Conostylideae), and Velloziaceae. Both Lachnanthes and Lophiola are included in this study. Schulze concluded that the Haemodoreae and Conostylideae are closely related on the basis of shared anatomical characteristics, predominantly the possession of stomatal subsidiary cells and a "mechanischer Ring" (sclerenchymatous sheath) in the underground stem. Lachnanthes differs from the remaining 6 genera of the Haemodoreae investigated in lacking axially extended epidermal cells which basally encircle the trichomes. Lophiola differs from the remaining 4 genera of the Conostylideae studied in lacking characteristic highly branched trichomes. More importantly, however, Lophiola stands apart from all other genera of the Haemodoreae and Conostylideae in lacking both stomatal subsidiary cells and the ring of sclerenchyma in the rhizome.

With the exception of Lachnanthes, all Haemodoraceae thus far examined have vessels confined to the roots, most of these with simple perforation plates (Cheadle 1968). Cheadle has noted, however, that three species of Tribonanthes, belonging to the tribe Conostylideae, and a single species of both Dilatris and Lanaria, belonging to the tribe Haemodoreae, possess root vessel elements with scalariform perforations. A similar condition is present in Lophiola. Although inconclusive in regards to the tribal relationships of Lophiola, these data tend to support the conjecture of Melchior (1964) and Cheadle (1969) that a natural alliance may exist between the Haemodoraceae and Tecophilaeaceae, which has vessel elements similar to Tribonanthes, Dilatris, Lanaria, and Lophiola. Lachnanthes, on the other hand, contains vessel elements with simple perforation plates in the root and ones with scalariform plates in the stem.

Within the Haemodorales (sensu Hutchinson 1934, 1959, 1973) Cheadle (1968) reported the Velloziaceae to have advanced vessel members in the roots and elements with scalariform perforations in stems and leaves. The Apostasiaceae is variable in vessel type and position, ranging from primitive elements that are restricted to roots to advanced vessel members in both root and stems. The Hypoxidaceae, Philydraceae, and Taccaceae all possess relatively primitive vessel elements in roots only.

Stebbins & Khush (1961) found all ten genera of *Haemodoraceae* examined by them to have stomata with two lateral subsidiary cells, generally confirming the results of Schulze (1893). Tomlinson (1974), furthermore, described the stomatal ontogeny of a single species of the haemodoraceous genus *Xiphidium* and showed that the subsidiary cells were derived from non-oblique divisions in a manner identical to that reported here for *Lachnanthes*. These two genera have consistently been placed in the same tribe (Geerinck 1969). The few specimens of *Hypoxidaceae*, *Velloziaceae*, and *Philydraceae* that have been studied also possess a pair of lateral subsidiary cells (Stebbins & Khush 1961). As far as is known, the *Taccaceae* lack subsidiary cells. The lack of subsidiary cells in *Lophiola*, accordingly, may prove taxonomically significant as additional members of the family are examined. Foliage of the putatively closely related family *Velloziaceae* differs noticeably from *Lachnanthes* and *Lophiola* in possessing prominent sclerenchymatous girders and strands and in lacking raphides (Ayensu 1969, 1974).

Despite the differences between Lophiola and Lachnanthes, the floral vasculature of both genera is basically similar to the general pattern described for the Liliaceae by Sterling (1972, 1977, and papers cited therein). In the lilies the floral venation is characterized by tepals being three trace structures, stamens receiving a single trace, and carpels being supplied by a dorsal bundle, 1—2 "septal" bundles, and 2 (4) placental bundles. Sterling (1972, 1973) tentatively described the following floral morphological features to be primitive in Liliaceae: incomplete fusion between carpel "wings", lack of fusion between pistil and floral cup, the presence of gynoecial sutural openings (most primitive) or sutural commissures, the presence of two septal bundles per carpel (as opposed to one or none), and the presence of bitegmic ovules.

Both Lachnanthes and Lophiola have laterally fused carpels with the ovary of Lophiola being half-inferior in position and that of Lachnanthes in a more advanced inferior position. Ovary position is correlated with the presence of carpellary sutural openings and stylar commissures in Lophiola and their absence in Lachnanthes. Both genera possess a single dorsal bundle per carpel and stylar canals. Advancement in the carpels of Lophiola is indicated by the presence of a solitary septal bundle whereas septal bundles are entirely absent in Lachnanthes. The occurrence of septal nectaries in Lachnanthes, a common feature in the Liliales (Brown 1938), undoubtedly represents an additional specialization. The presence of 2 to 4 placental bundles in both genera is similar to the pattern of ovular vascularization in the Liliaceae. In contrast to several species of the primitive liliaceous subfamily Wurmbaeoideae (Sterling 1972), both Lachnanthes and Lophiola lack gynoecial vascular bundles that arise laterally from the dorsal carpellary bundle.

Although it is abundantly clear that considerably more detailed and extensive information is required in order to satisfactorily assess the evolutionary relationships of the Haemodoraceae and related families, available anatomical and morphological data tends to refute the placement of Lophiola in the tribe Haemodoreae as done by Geerinck (1969). The same conclusion was reached by Robertson (1976). Lophiola demonstrates several anatomical dissimilarities, both from Lachnanthes, as seen in the present study, and, as recorded by Schulze (1893), from other members of both the Haemodoreae and Conostylideae. ERDTMAN (1966) observed that Lophiola differs palynologically from all other investigated taxa of the Conostylideae (and Haemodoreae) and is perhaps more closely related to the Conanthereae (= Tecophilaeaceae, sensu Hutchinson 1934, 1959, 1973). This conjecture is in general agreement with that of Cheadle (1969), based on vessel element morphology. However, Orn-DUFF (1979), based on chromosome counts of several genera of the Haemodoraceae, including Lachnanthes and Lophiola, concluded that Lophiola "seems more at home in the Haemodoreae than in the Constylideae". Additional study of the family appears necessary before the taxonomic placement of Lophiola can be firmly established.

A continuing investigation of the Haemodoreae is underway by the senior author.

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